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Thromboxane A2 Receptor Blocker, Seratrodast: A Novel Controller Medication For The Management Of Asthma

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Introduction:

Asthma is a chronic inflammatory disease of the airways that is characterized by widespread obstruction of the airways, usually reversible, and bronchial hyper-responsiveness. As per GINA, Japanese and ICS¹ guidelines for management of asthma, medications to treat asthma can be classified as relievers or controllers. Relievers are medications used on an as-needed basis that act quickly to reverse bronchoconstriction and relieve its symptoms^{1,2}. They include rapid-acting inhaled β_2 -agonists, inhaled anticholinergics, short-acting theophylline, and short-acting oral β_2 -agonists. Controllers are medications taken daily on a long-term basis to keep asthma under clinical control chiefly through their anti-inflammatory effects^{1,2}. They include inhaled and systemic glucocorticosteroids, leukotriene modifiers, long-acting β_2 -agonists, sustained-release theophylline, cromones and anti-IgE.

"Inadequate control of asthma" was determined to be the common cause of the results obtained in the afore mentioned surveys, which was corroborated by other studies wherein patients had uncontrolled asthma despite guideline-based use of inhaled corticosteroids (ICS) and inhaled long-acting β_2 -agonists (LABAs)³. With respect to rise in uncontrolled asthma, the 2002 GINA guidelines stated: "it is reasonable to expect that in most patients with asthma, control of the disease can and should be achieved and maintained"¹. To meet this challenge, the recent GINA guidelines of year 2011, not only incorporates updated scientific information but also describes a development of the aforementioned theme and a change in approach to asthma management, with asthma control, rather than asthma severity, being the focus of treatment decisions. The GINA guidelines define control of asthma as minimal chronic symptoms, minimal (infrequent) exacerbations, no emergency visits, minimal use of as-needed (rapid-acting) β_2 -agonists, no limitations on activities, daily peak expiratory flow (PEF) variation of less than 20%, near normal PEF and minimal adverse effects from medications^{1,2}. In the stepwise approach to therapy recommended in the GINA guidelines, a reliever medication (rapid-onset bronchodilator, either short-acting or long-acting) should be provided for quick relief of symptoms. However, regular use of reliever medication is one of the elements defining uncontrolled asthma, and indicates controller treatment should be increased.⁴

Thus reducing or eliminating the need for reliever treatment is an important GINA goal and a measure of success of treatment, which is achieved by long-term management of asthma with controller agents^{1,2}. Controllers alleviate and eliminate asthma symptoms, and normalize and maintain the conditions. As mentioned earlier they include inhaled and oral corticosteroids, long-acting β_2 agonists (LABA), leukotriene receptor antagonists (LTRAs), theophylline, anti-IgE antibody, H1 receptor antagonists, Th2 cytokine inhibitor and last but not the least thromboxane A2 modulators.

Thromboxane A2 & Thromboxane A2 Receptor in the Pathogenesis of Asthma

Thromboxane A2 (TXA₂), a cyclooxygenase product of arachidonic acid, has been implicated in the

pathogenesis of asthma. The recent demonstration that in man all the bronchoconstrictor prostanoids exert their effects by binding to the TXA₂ receptor (TP receptor) has aroused a renewed interest in the role of TXA₂ and TP receptor-related prostaglandins in asthma. In addition, the recent development of specific and potent TP receptor antagonists and inhibitors of thromboxane synthase (TX synthase) has provided tools to assess the role of these mediators in the pathophysiology of asthma⁵². Both GINA^{10,11} and Japanese guidelines for management of asthma⁹⁵ have indicated the use of TXA₂ modulators as controllers as step 1 for long-term management of asthma.

Cellular origin of TXA₂

TXA₂ is synthesized by enzyme TX synthase, which has been found to be associated with alveolar macrophages^{5,6}, bronchial epithelial cells and human lung fibroblasts. In addition to alveolar macrophages, TXA₂ is the major product of the cyclooxygenase pathway in three types of human blood cells: platelets^{7,8}, eosinophils⁹ and monocytes^{10,11}. The eosinophils, which infiltrate the airway wall in asthma, are also an important source of TXA₂ since human blood eosinophils obtained from asthmatics generate and release much more TXA₂ than the other cyclooxygenase metabolites under resting conditions or after activation by PAF¹². In addition, TXA₂ originated from platelets may contribute to lung inflammation in asthma. Close association between platelets, eosinophils and monocytes or alveolar macrophages has been observed causing increase in TXA₂ production¹³.

Heterogeneity of TP receptors

Prostanoid TP receptors are distributed in both plasma membrane and cytosolic compartments. They are found in tissues rich in vasculature such as lung, heart, and kidney. Two G protein-coupled isoforms of this receptor have been subsequently described, TP α (placental/platelet) and TP β (endothelial) that differ in length and sequence of the carboxyl-terminal tail distal to Arg328. Intra-receptor differences in C-terminal tail sequence allows for significant differences in their ability to internalize in response to agonist exposure¹⁴. For eg. TXA₂ mimetic (analogue) was observed to bind to TP β and not TP α .

Biological activities of TXA₂ and related PGs in airways

1. Bronchial Smooth muscle contraction

Ligand binding studies on TP receptors have shown their existence on airway smooth muscle. Comparisons of the in vitro contractile potencies of TXA₂ mimetic, PGF₂ α , PGD₂ and 9 α ,11 β -PGF₂ on airway smooth muscle have shown that the TXA₂ analogue is about two orders of magnitude more potent than the other PGs¹⁵⁻¹⁷. Experiments utilizing several TXA₂ receptor antagonists have further supported the view that not only the TXA₂ mimetic but also PGF₂ α , PGD₂ and its metabolite 9 α ,11 β -PGF₂ contract bronchial smooth muscle by acting via direct stimulation of TP receptors. Thus, TXA₂ and other prostanoids play an important role in airway smooth muscle contractions.

2. Vascular smooth muscle contraction

TXA₂ is one of the most potent constrictors of human vascular smooth muscle as illustrated by its in vitro potency and efficacy on internal mammary arteries or saphenous veins^{18,19}. The picture appears more complex than for the airway smooth muscle because different prostanoid receptors are involved in the control of the vascular tone and because the prostanoid effects vary with species, vascular sites and age. However, activation of vascular TP receptors invariably induces vasoconstriction.

3. Plasma Extravasation

TXA₂ mimetic and with less potency, PGF₂ α , were found to potently induce plasma exudation in airways²⁰. Plasma exudation induced by inhaled PAF or by leukotriene D₄ (LTD₄) instilled via the airway route was observed to be partly inhibited by TX synthase inhibitor and TP receptor antagonists, suggesting that TXA₂ is endogenously released in response to inflammatory mediators other than prostanoids resulting in airway microvascular leakage.

4. Neuromodulatory effects

Prostanoids have potent neuromodulatory effects. Thromboxane receptor agonists were observed to elevate electrically induced norepinephrine release in an isolated arterial preparation, suggesting that TXA2 and related prostaglandins may have some vascular effects on adrenergic fibres that have been found to be in close association with bronchial vessels²¹. TXA2 has also been implicated in acting presynaptically to enhance the release or duration of release of acetylcholine, a potent bronchoconstrictor, from cholinergic nerves²¹.

5. Mucous secretion

In human airway tissue implants, PGF₂ and PGD₂ significantly increased mucous glycoprotein release^{22,23}. In addition, TXA2 was shown to increase tracheal mucous gel layer, which was inhibited by TP receptor antagonist. In the same study, the TP receptor antagonist was observed to attenuate mucous gel layer response caused by leukotrienes, indicating an indirect link between leukotrienes and TP receptors²⁴.

6. Smooth muscle proliferation

TXA2 elicits the proliferation of human airway smooth muscle cells as well as vascular smooth muscle cells^{25,26}. It may therefore participate in airway remodeling in asthma that includes airway smooth muscle hypertrophy and hyperplasia²⁷.

7. Airway hyperresponsiveness

The effects of TXA2 and related PGs on plasma exudation, acetylcholine release and smooth muscle proliferation support the potential role of TP receptor stimulation in the pathogenesis of airway hyperresponsiveness²⁸. The airway mucosal edema due to plasma exudation and the smooth muscle proliferation contribute to thickening of the airway wall. In addition, plasma exudation may lead to liquid filling of the airway interstices formed between luminal epithelial projections. This liquid filling of airway interstices amplify the luminal narrowing due to airway smooth muscle contraction. The enhancing effect of TXA2 on cholinergic transmission is also a possible mechanism by which TXA2 may cause airway hyperresponsiveness²⁸.

Various thromboxane synthase inhibitors have been shown to prevent increased airway reactivity to allergens²⁹, PAF (30), LTC₄, D₄^{31,32} and B₄³³, bradykinin³⁴, endothelin³⁵, endotoxin³⁶ and ozone³⁷. It has been suggested that these mediators induce airway hyperresponsiveness through the generation of TXA₂, in parallel with other effects on the airway smooth muscle³⁸.

8. TXA₂ in COPD

TXA₂ is also involved in pathogenesis of COPD. It was demonstrated in patients with COPD that urinary excretion of 11-dehydro-TXB₂, a TXA₂ metabolite, was significantly higher than in healthy subjects, suggesting an enhancement of platelet TXA₂ biosynthesis in such patients³⁹. Thromboembolic events as pulmonary hypertension are complications of COPD and TXA₂ may play an important role in these complications. It was shown that TXA₂ level is significantly increased in patients suffering from COPD with pulmonary hypertension in comparison with that of normal subjects, suggesting TXA₂ plays an important role in causing pulmonary hypertension⁴⁰.

The above biological activities of thromboxane A₂ indicate that TXA₂ is involved in various steps in progression of asthma. More importantly the direct action of prostanoids (TXA₂, PGF₂α, PGD₂ and its metabolite 9α,11β-PGF₂) via activation of TP receptors and the indirect action of leukotrienes and other agents on TP receptors through intricate pathways, give a new insight into the mechanisms underlying pathogenesis of asthma and suggest the importance of TP receptor modulation for management of asthma.

Pharmacology Of Txa2 Modulators In Asthma

Strategies for inhibition of TXA2 include inhibition of thromboxane synthase and antagonism of TP receptors. TX synthase inhibitors effectively suppress TXA2 synthesis; however, inhibition of TX synthase results in an increased availability of the precursor PGH2 with the possibility of prostanoid synthesis redirection from TXA2 to other prostaglandins according to the enzyme equipment of the cells⁴¹.

Therefore, accumulated PGH2 or increased synthesis of prostaglandins, such as PGF2 α and PGD2, may substitute for TXA2 in causing TP receptor stimulation or may increase the stimulation of other prostanoid receptors with a resulting reduction in the potential effects of TX synthase inhibition⁴¹.

Bleeding tendency is one of the possible adverse effects of anti-TXA2 agents due to suppression of platelet aggregation. This is much more pronounced with TX synthase inhibitors because PGI2 production, which has anti-platelet aggregation activity, is enhanced by pathway blockade from PGH2 to TXA2 by TX synthase inhibitors⁴².

Thromboxane A2 receptor (TP receptor) antagonism

As indicated earlier, the second way of modulation of TXA2 is by antagonizing TP receptors, which is better and safer than TX synthase inhibition. TP receptor antagonists not only inhibit the actions of TXA2, but also inhibit the activation of TP receptors by other prostanoids (PGF2 α , PGD2 and its metabolite 9 α ,11 β -PGF2). Additionally, TP receptors antagonists also act on the intricate pathways involved in activation of TP receptors by allergens, PAF, leukotrienes, bradykinin, endotoxin, endothelin and ozone.

Seratrodast: It is an orally active quinone derivative and a potent TP receptor antagonist⁴³⁻⁴⁷. It was the first thromboxane receptor antagonist that was developed as an anti-asthmatic drug and received marketing approval in Japan in 1997, and has been in use from past 15 years as a controller therapy in the management of asthma.

Seratrodast has been shown to inhibit bronchoconstriction induced by TXA2, PGD2, 9 α ,11 β -PGF2, PGF2 α , leukotriene D4 (LTD4) and PAF^{43, 44, 48}, and has been observed to affect both immediate and late asthmatic responses^{47, 49}. Seratrodast has been reported to reduce airway hyper-responsiveness to various stimuli^{47, 49-51}. Seratrodast abolished the decrease in internal diameter of pulmonary arteries caused by vagal nerve stimulation or injection of acetylcholine⁴⁸. Seratrodast administered orally at doses of 1-10 mg/kg 1 h before the challenge with antibody clearly inhibited the pulmonary pressure increase caused by Forssman anaphylaxis. At a dose of 10 mg/kg, seratrodast also inhibited the pulmonary pressure increase caused by LTD4 and TXA2 mimetic⁵².

Pharmacokinetic profile of Seratrodast in plasma was determined in an open labeled, two treatment, two period, two sequence, single dose, comparative, randomized, two way crossover design under non fed condition with 14 days washout period in 26 healthy male subjects⁵³. Seratrodast showed a long half of 22 hours with an approximate of 3.5 hours to attain maximal plasma concentration. Clinical studies of Seratrodast have been conducted on approximately 5000 patients (post-marketing surveillance study on 3000 patients and clinical studies in approximately 2000 patients) in various indications like asthma, chronic bronchitis, chronic pulmonary emphysema, perennial allergic rhinitis, and chronic cough (table-1).

Table 1: Clinical studies on Seratrodast

SN	Test drug	Study Design	Duration of the study	Patients	Conclusion
1	Seratrodast 80 mg Vs Montelukast 10 mg	Double-blind, double dummy Randomized	4 weeks	205 Indian patients with Asthma on ICS therapy	1. Significant improvement in FEV1, FVC and PEF; and in clinical symptoms of asthma such as Wheezing, shortness of breath, expectoration and chest tightness were observed 2. Greater reduction in sputum parameters such as levels fucose, ECP, albumin and expectoration as compared to Montelukast (54)
2	Seratrodast 80mg Montelukast 10mg	Double-blind, randomized	8 weeks	213 Asthma patients	Seratrodast is safe, and effective drug in treating patients with asthma and possesses reliable efficacy and safety similar to montelukast. (55)
3	Seratrodast 40 Zafirlukast 20	Double-blind, randomized, Multi centric	6 weeks	220 Asthma patients	The control rate of seratrodast (71.68%) was significantly higher than that of zafirlukast (62.62%). (56)
4	Seratrodast 80mg od, Placebo, Pranlukast 112.5 mg t.i.d.	Randomized cross-over trial	4 weeks	16 Chronic Bronchitis patients	The cough threshold was significantly increased compared with placebo after four-week treatment with seratrodast, but not after treatment with pranlukast. Thromboxane antagonism may be considered to be one of the therapeutic options for the treatment of chronic productive cough. (57)
5	Seratrodast 80mg OD	Open-label, crossover design study	12 weeks	10 Asthma patients	i. Seratrodast decreased the concentration of ECP in sputum significantly ($p < 0.05$). ii. Seratrodast improves clinical symptoms and airway hyperresponsiveness by reducing airway inflammation. (58)
6	Seratrodast 80mg o.d.	Open label study	8 weeks	14 Chronic pulmonary Emphysema patients	i. The results revealed significant improvement of respiratory distress, evaluated on both the Hugh-Jones classification and the Borg scale, at week 8. ii. Among the respiratory function parameters examined, only FVC was significantly improved. (59)
7	Seratrodast 80mg o.d.		24 months	35 Asthma patients	The exacerbation rate in patients taking seratrodast after 12 months tended to be lower than in the other two patient groups ($P = 0.0743$). In seratrodast group, the average dose of iBDP from 9 to 24 months after the initial step down was significantly lower than before the step down ($P < 0.0001$) and was not significantly different from the mean dose of iBDP in groups A or B. (60)

SN	Test drug	Study Design	Duration of the study	Patients	Conclusion
8	Seratrodast 40mg o.d., Placebo	Double blind, randomized	6 weeks	45 Asthma patients	<p>i. Seratrodast treatment decreased the amount of sputum ($p = 0.005$), dynamic viscosity ($p = 0.007$), and albumin concentration ($p = 0.028$).</p> <p>ii. Nasal clearance time of a saccharin particle was shortened in the seratrodast group at week 4 ($p = 0.031$) and week 6 ($p = 0.025$), compared with the placebo group. (61)</p>
9	Seratrodast 80mg/d Terfenadine 120 mg/d	Double-blind Multi-center	-	234 Allergic Rhinitis patients.	The improvement rate of nasal obstruction nasal discharge and sneezing at the final evaluation in the seratrodast group was significantly superior to Terfenadine group, 51.4% vs. 34.7% ($p=0.016$). (62)
10	Seratrodast 40mg o.d.	Open label study	4 weeks	45 Asthma patients	The ratio of urinary eicosanoids might be a possible predictor of the effects of TXA2 receptor antagonist. (63)
11	Seratrodast 80mg o.d., Placebo	Double blind, randomized,	4 months	31 Asthma patients	Seratrodast in asthma may inhibit activated eosinophil infiltration in part by modulating the expression of chemokines in bronchial tissues. (64)
12	Seratrodast 80mg/d or 120mg/day	Double blind, randomized	8 weeks	183 asthma patients	<p>i. Seratrodast at a dose of 120 mg daily produced an increase in forced expiratory volume in 1 second (FEV1) from baseline that was linearly correlated with its plasma concentrations</p> <p>ii. A lower percentage of predicted FEV1 (i.e., more severe obstruction) was associated with higher slopes, and greater increases in FEV1. (65)</p>
13	Seratrodast 20/40/80mg/d	Randomized,	4 weeks	155 allergic rhinitis patients	Improvement rates for nasal obstruction, nasal discharge and sneezing showing "marked improvement" or better in the 20mg, 40mg, and 80mg daily doses. (66)
14	Seratrodast 4mg/day, 40mg/day or 80mg/day	Multi-center, Double-blind Comparative Study	4 weeks	224 Allergic Rhinitis	Improvement rate evaluation of nasal symptoms, was dose-dependently. In the subject group whose main complaint was nasal obstruction & duration of disease was 1 year or longer, improvement rate in nasal symptoms (final evaluation) was similarly dose-proportional. (67)
15	Seratrodast 80mg/day	Open label study	12-24 weeks	19 Allergic Rhinitis	Seratrodast appears to be highly effective in treating perennial allergic rhinitis and safe in a long-term period. (68)

SN	Test drug	Study Design	Duration of the study	Patients	Conclusion
16	Seratrodast 20 or 80mg/day		4 weeks	38 Allergic Rhinitis	1) In the 80mg/day group, changing rates of nasal volume and minimum cross section showed significant improvement after 4 weeks. 2) Improvement rate in nasal obstruction in the 80mg/day group was higher than that in the 20mg/day group. 69

Discussion

TXA2 is an important mediator in the regulation of eosinophil degranulation⁷⁰. TXA2 increases eosinophil degranulation resulting in greater ECP release. Seratrodast decreased (ECP) concentrations in sputum of asthma patients⁵⁴. The decrease in ECP levels with Seratrodast was significantly greater than that observed with Montelukast (-27.20 vs. -23.55 ng/ml, $p < 0.05$) indicating dominance of TXA2 over leukotrienes in modulating eosinophil degranulation (54). This implies that TP receptor antagonism may have a better impact than cyteinyl leukotriene receptor antagonism over severity of asthma.

Effect on airway microvascular leakage

In patients with asthma, plasma-protein leakage in the airway appears to correlate with indirect indexes of airway inflammation⁷¹. Being a major plasma protein, albumin level in sputum has been suggested to be a biochemical marker for microvascular leakage⁷². In a 4-week comparative clinical trial conducted in 205 Indian patients⁵⁴, significantly greater reduction in sputum albumin levels were observed with Seratrodast compared to Montelukast (-37.51 vs. -32.82 mg/dl, $p < 0.05$).

Effect on dynamic viscosity and elasticity of sputum

In various studies, sputum fucose levels have been used as biochemical marker for viscosity and elasticity of respiratory mucus⁷³. Fucose, a respiratory mucus glycoprotein, is produced by goblet cells and submucosal gland cells in the respiratory mucosa⁷⁴, and largely contributes to viscoelasticity of nasal mucus. The relative levels of fucose in respiratory mucus depend on degree of hypertrophy and hyperplasia of airway smooth muscle, which is increased by TXA2. Seratrodast, by TP receptor antagonism, inhibits the influence of TXA2 on airway smooth muscle which in turn affects the level of fucose in respiratory mucus. Seratrodast has been observed to decrease the levels of sputum fucose^{54, 61} resulting decrease in dynamic viscosity and elasticity of sputum.

Effect on peak expiratory flow (PEF) and other lung function parameters

Prostanoids are the major inflammatory mediators involved in airway smooth muscle contraction/bronchoconstriction. Various pre-clinical studies have indicated predominant role of prostanoids in airway smooth muscle contraction compared to that of leukotrienes⁷⁵. Seratrodast was observed to have a radical impact on PEF in clinical trial conducted on Indian patients⁵⁴. A mean change of 0.614 L/sec from baseline was observed with Seratrodast in comparison to 0.199 L/sec with Montelukast, equivalent to 24.9L/min higher PEF with seratrodast ($P < 0.05$).

In addition to PEF, significant improvements in other lung function parameters (FVC and FEV1) from baseline have been observed with Seratrodast in various clinical studies^{54, 59, 62, 66, 69, 76}, indicating the efficacy of TP blockers in alleviating bronchoconstriction.

Effect on amount of mucous secretion

TXA2, and other prostanoids PGF2? & PGD2, significantly increases mucous glycoprotein release and

tracheal mucus gel layer²⁴ that indicates increase in sputum secretion. In various clinical studies, Seratrodast has been observed to limit the sputum secretion action of prostanoids^{61, 62, 66, 76}. In a clinical trial comparing the efficacy of Seratrodast with Montelukast, significantly greater reduction in expectoration was observed with Seratrodast than with Montelukast⁵⁴.

Effect on airway hyper-responsiveness

In a 12-week study in 10 asthma patients⁵⁷, Seratrodast improved airway hyper-responsiveness by reducing airway inflammation indicating close association of TP receptor antagonism with improvement in airway hyper-responsiveness. TP receptor antagonism results in direct inhibition of actions of prostanoids on TP receptors and indirect inhibition of effect of allergens, leukotrienes, bradykinin, PAF, endothelin, endotoxin and ozone on TP receptors^{47; 49-51}. The decrease in airway hyper-responsiveness by Seratrodast can also be correlated with reduction in ECP that contributes to airway hyper-reactivity⁷⁷.

Effect on cough threshold

Seratrodast was observed to increase cough threshold in patients with chronic bronchitis, indicating involvement of prostanoids in chronic cough. Previous *in vitro* studies have shown the involvement of cyclooxygenase products of arachidonic acid metabolism, especially TXA₂, to be involved in increased mucus secretion in respiratory tract⁴¹, which in turn stimulates mechanoreceptors causing cough. In clinical study conducted by Ishiura et al⁵⁶ in patients with chronic bronchitis, Seratrodast, after 4 weeks of treatment, was observed to increase cough threshold indicating that inflammatory mediators such as prostanoids can adjust the sensitivity of cough reflex despite the direct stimulation of mechanoreceptors by excessive mucus in chronic bronchitis. Pranlukast, on the other hand, did not increase cough threshold, implying that cysteinyl leukotriene is probably not involved in modulation of airway cough sensitivity.

TP receptor antagonists and Cysteinyl leukotriene receptor antagonists

There are various inflammatory pathways involved in the pathogenesis of asthma that involve secondary TXA₂ synthesis by multiple agonists including leukotrienes⁷⁸. This is corroborated by various studies wherein inhibiting TXA₂ prevented increased airway reactivity to allergens²⁹, PAF³⁰, LTC₄, D₄^{31,32} and B₄³³, bradykinin³⁴, endothelin³⁵, endotoxin³⁶ and ozone³⁷.

Both GINA^{1,2} and Japanese asthma guidelines⁷⁹ recommend the use of cysteinyl leukotriene receptor antagonists and TP receptor antagonists for managing uncontrolled asthma. Considering the role of TXA₂ in pathogenesis of asthma and the efficacy of TP receptor antagonists over cysteinyl leukotriene receptor antagonists, TP receptor antagonism could soon become a new paradigm in management of uncontrolled asthma.

Conclusion

Since the identification and isolation of the first prostaglandins in the 1960s, the cyclooxygenase product (prostanoids) of arachidonic acid have been implicated in asthma⁴¹. TXA₂ has been implicated in airway smooth muscle contraction, vascular smooth muscle contraction, airway microvascular leakage, bronchoconstriction through neuromodulatory effects, increased nasal mucous secretion, airway remodeling and airway hyper-responsiveness⁴¹. All bronchoconstrictor prostanoids i.e. PGD₂, PGF₂ and TXA₂, act via thromboxane A₂ receptors (TP receptors), that are distributed in both plasma membrane and cytosolic compartments found in tissues rich in vasculature such as lung, heart, and kidney⁸⁰. Seratrodast effectively improves nasal obstruction, lung function parameters, cough threshold⁵⁶, airway hyper-responsiveness⁵⁷ and mucociliary clearance⁵⁴, and helps in reduction of average dose of inhaled corticosteroids⁶¹. Mucous secretion and sputum viscosity are significantly decreased by Seratrodast^{54,61}. Clinical symptoms of asthma, like wheezing, breathlessness, expectoration, cough and chest tightness are significantly improved⁵⁴. A better control over exacerbation rate was observed with

Seratrodist in comparison to cysteinyl leukotriene receptor antagonists like Pranlukast, Zafirlukast and Montelukast⁵⁶. Both GINA guidelines and Japanese guidelines for management of asthma recommend the use of Seratrodist in uncontrolled asthma. With the implication of involvement of TXA2 in pulmonary hypertension in COPD patients and increase in levels of TXA2 during COPD⁴⁰, respiratory distress syndrome¹⁴ and chronic pulmonary emphysema⁵⁹, Seratrodist could be effective in these respiratory disorders other than asthma. The first Indian study conducted by Zuventus Healthcare Ltd., on Seratrodist in patients with mild to moderate asthma indicated better reduction of sputum ECP, sputum albumin and expectoration with Seratrodist in comparison to that of Montelukast⁵⁴. Seratrodist showed significantly greater improvement in PEF compared to Montelukast⁵⁴, indicating better control over asthma than Montelukast. Further evidence from clinical studies and clinical usage in Indian patients is suggested to firm up the role of seratrodist in patients with asthma, allergic rhinitis and COPD.

References

1. Global Initiative for Asthma. Global strategy for asthma management and prevention. NIH publication no. 02-3659. 2002. National Institutes of Health/National Heart, Lung, & Blood Institute
2. Global Initiative for Asthma. Global strategy for asthma management and prevention. NIH publication no. 02-3659 (updated 2004). 2004. National Institutes of Health/National Heart, Lung, and Blood Institute
3. Peters SP, Ferguson G, Deniz Y, Reisner C. Uncontrolled asthma: a review of the prevalence, disease burden and options for treatment. *Respir Med.* 2006 Jul;100(7):1139-51
4. Chung KF, Godard P, Adelroth E, et al. *Eur Respir J* 1999;13:1198-208
5. Robinson C, Hardy CC, Holgate ST. Pulmonary synthesis, release and metabolism of prostaglandins. *J Allergy Clin Immunol* 1985;76:265-71
6. Balter MS, Eschenbacher WL, Peters-Golden M. Arachidonic acid metabolism in cultured alveolar macrophages from normal, atopic and asthmatic Subjects. *Am Rev Respir Dis* 1988;138: 1134-42
7. Gresele P, Deckmyn H, Nenci GG, Verrnylen J. Thromboxane synthase inhibitors, thromboxane receptor antagonists and dual blockers in thrombotic disorders. *Trends Pharmacol Sci* 1991;12:158-63
8. Hamberg M, Svensson I, Samuelsson B. Thromboxanes. A new group of biologically active compounds derived from prostaglandin endoperoxydes. *Proc Natl Acad Sci USA* 1975;72:2994-8
9. Foegh ML, Maddox YT, Ramwell PW. Human peritoneal eosinophils and formation of arachidonate cyclooxygenase products. *Scand J Immunol* 1982;23:599-603
10. Jones CM, Hall ER, Hester JP, Wu KK. Arachidonic acid metabolites produced by platelet-depleted human blood monocytes: a possible role in thrombogenesis. *Am J Hematol* 1989;31:145-52
11. Juergens UR, Christiansen SC, Stevenson DD, Zuraw BL. Arachidonic acid metabolism in monocytes of aspirin-sensitive asthmatic patients before and after oral aspirin challenge. *J Allergy Clin Immunol* 1992;90:636-45
12. Kroegel C, Matthys H. Platelet-activating factor-induced human eosinophil activation. Generation and release of cyclo-oxygenase metabolites in human blood eosinophils from asthmatics. *Immunology* 1993;78:279-85
13. Maghni K, Carrier J, Cloutier S, Sirois P. Cell-cell interactions between platelets, macrophages, eosinophils and natural killer cells in thromboxane A2 biosynthesis. *J Lipid Mediat* 1993;6:321-32

14. Deby-Dupont G, Braun M, Lamy M, Deby C, Pincemail J, Faymonville ME, Damas P, Bodson L, Lecart MP, Goutier R. Thromboxane and prostacyclin release in adult respiratory distress syndrome. *Intensive Care Med.* 1987;13(3):167-74
15. Beasley R, Varley I, Robinson C, Holgate ST. Cholinergic mediated bronchoconstriction induced by prostaglandin (PG)D₂ and its initial metabolite 9 β ,11 β -PGF₂ and PGF₂ α in asthma. *Am Rev Respir Dis* 1987a; 136: 1140-4
16. Coleman RA, Sheldrick RLG. Prostanoid-induced contraction of human bronchial smooth muscle is mediated by TP receptors. *Br J Pharmacol* 1989;90:688-92
17. Featherstone RL, Robinson C, Holgate ST, Church MK. Evidence for thromboxane receptor mediated contraction of guinea-pig and human airways in vitro by prostaglandin (PG)D₂, 9 β ,11 β -PGF₂ and PGF₂ α . *Naunyn Schmiedebergs Arch Pharmacol* 1990;341:439-41
18. He GW, Rosenfeldt FL, Buxton BF, Angus JA. Reactivity of human isolated internal mammary artery to constrictor and dilator agents. *Circulation* 1989;80(suppl. I):I-141-50
19. Schilling A, Glusa E, Miiller-Schweintzer E. Nature of the vehicle solution for cryopreservation of human peripheral veins. Preservation of reactivity to pharmacological stimuli. *Cryobiology* 1995;32: 109-13
20. Lotvall J, Elwood W, Tokuyama K, Sakamoto T, Barnes PJ, Chung KF. A thromboxane mimetic, U-46619, produces plasma exudation in airways of the guinea pig. *J Appl Physiol* 1992; 72:2415-9
21. Barnes PJ. Neural control of human airways in health and disease. *Am Rev Respir Dis* 1986; 134: 1289-314
22. Marom Z, Shelhamer JH, Kaliner M. The effects of arachidonic acid, monohydroxyeicosatetraenoic acid and prostaglandins on the release of mucous glycoproteins from human airways in vitro. *J Clin Invest* 1981 ;67:1695-703
23. Rich B, Peatfield AC, Williams IP, Richardson PS. Effects of prostaglandins E₁, E₂ and F₂ α on mucin secretion from human bronchi in vitro. *Thorax* 1984;39:420-3
24. Yanni JM, Smith WL, Foxwell MH. U46619 and carboxylic thromboxane A₂-induced increases in tracheal mucous gel layer thickness. *Prostaglandins Leukotrienes Essent Fatty Acids* 1988;32:45-9
25. Morinelli TA, Zhang LM, Newman WH, Meier KE. Thromboxane A₂/Prostaglandin H₂-stimulated mitogenesis of coronary artery smooth muscle cells involves activation of mitogen-activated protein kinase and S6 kinase. *J Biol Chem* 1994;269:5693-6
26. Tomlinson PR, Wilson JW, Stewart AG. Inhibition by salbutamol of the proliferation of human airway smooth muscle cells grown in culture. *Br J Pharmacol* 1994;111:641-7
27. Stewart AG, Tomlinson PR, Wilson J. Airway wall remodeling in asthma: a novel target for the development of antiasthma drugs. *Trends Pharmacol Sci* 1993;14:275-9
28. O'Byrne PM, Fuller RW. The role of thromboxane A₂ in the pathogenesis of airway hyperresponsiveness. *Eur Resp J* 1989;2:782-6
29. Minoguchi K, Adachi M, Tokunaga H, et al. Change in responsiveness of airway and beta-adrenoceptor in guinea pigs. *Amerugi* 1993; 42 (4): 556-63
30. Chung KF, Aizawa H, Leikauf GD, et al. Airway hyperresponsiveness induced by platelet-activating factor: role of thromboxane generation. *J Pharmacol Exp Ther* 1986; 236 (3): 580-4
31. Muccitelli RM, Osborn RR, Weichman BM. Effect of inhibition of thromboxane production on the leukotriene D₄-mediated bronchoconstriction in the guinea pig. *Prostaglandins* 1983;26 (2):197-206

32. Kurosawa M, Tsukagoshi H. Inhibitory effect of a thromboxane A2 synthetase inhibitor OKY-046 on bronchial hyperresponsiveness to histamine, but not on airway wall thickening, induced by intravenous administration of leukotriene C4 in guinea-pigs. *Pulm Pharmacol* 1993; 6 (4): 247-53
33. O'Byrne PM, Leikauf GD, Aizawa H, et al. Leukotriene B4 induces airway hyperresponsiveness in dogs. *J Appl Physiol* 1985; 59 (6): 1941-6
34. Ueno A, Tanaka K, Katori M. Possible involvement of thromboxane in bronchoconstrictive and hypertensive effects of LTC4 and LTD4 in guinea pigs. *Prostaglandins* 1982; 23 (6): 865-80
35. Ninomiya H, Yu XY, Hasegawa S, et al. Endothelin-1 induces stimulation of prostaglandin synthesis in cells obtained from canine airways by bronchoalveolar lavage. *Prostaglandins* 1992; 43 (5): 401-11
36. Held HD, Uhlig S. Mechanisms of endotoxin-induced airway and pulmonary vascular hyperreactivity in mice. *Am J Respir Crit Care Med* 2000; 162 (4 Pt 1): 1547-52
37. Aizawa H, Chung KF, Leikauf GD, et al. Significance of thromboxane generation in ozone-induced airway hyperresponsiveness in dogs. *J Appl Physiol* 1985; 59 (6): 1918-23
38. Kurosawa M. Role of thromboxane A2 synthetase inhibitors in the treatment of patients with bronchial asthma. *Clin Ther* 1995; 17 (1): 2-11
39. Davi, G., Basili, S., Vieri, M., Cipollone, F., Santarone, S., Alessandri, C., Gazzaniga, P., Cordova, C., Violi, F., 1997. Enhanced thromboxane biosynthesis in patients with chronic obstructive pulmonary disease. The Chronic Obstructive Bronchitis and Haemostasis Study Group. *Am. J. Respir. Crit. Care Med.* 156, 1794-99
40. Zheng, L., Duan, S.F., Zhang, Z.X., 1991. Changes of thromboxane A2 (TXA2) and prostacyclin (PGI2) in COPD patients with pulmonary hypertension. *Zhonghua Nei Ke Za Zhi* 30, 91-93, 126
41. Devillier P, Bessard G. Thromboxane A2 and related prostaglandins in airways. *Fundam Clin Pharmacol.* 1997;11(1):2-18
42. Mizushima Y, Oosaki R. Clinical Potential of Anti-Thromboxane A2 Agents in Bronchial Asthma. *BioDrugs.* 1997 Feb;7(2):91-8
43. Ashida Y, Matsumoto T, Kuriki H, et al. A novel anti-asthmatic quinone derivative, AA-2414 with a potent antagonistic activity against a variety of spasmogenic prostanoids. *Prostaglandins* 1989;38(1):91-112
44. Shiraishi M, Kato K, Terao S, et al. Quinones. 4. Novel eicosanoid antagonists: synthesis and pharmacological evaluation. *J Med Chem* 1989; 32 (9): 2214-21
45. Fukumoto S, Shiraishi M, Terashita Z, et al. Synthesis and thromboxane A2/prostaglandin H2 receptor antagonistic activity of phenol derivatives. *J Med Chem* 1992; 35 (12): 2202-9
46. Kurokawa T, Matsumoto T, Ashida Y, et al. Antagonism of the human thromboxane A2 receptor by an anti-asthmatic agent AA-2414. *Biol Pharm Bull* 1994;17(3):383-5
47. Terao S, Shiraishi M, Matsumoto T, et al. Thromboxane A2 antagonist -discovery of seratrodist *Yakugaku Zasshi* 1999; 119 (5): 377-90
48. Samara EE. Seratrodist (AA-2414)-A Novel Thromboxane-A2 Receptor Antagonist. *Cardiovasc Drug Rev* 1996; 14(3):272-85
49. Matsumoto T, Ashida Y, Tsukuda R. Pharmacological modulation of immediate and late airway response and leukocyte infiltration in the guinea pig. *J Pharmacol Exp Ther* 1994; 269 (3): 1236-44
50. Imai T, Adachi M, Idaira K, et al. The effect of a specific thromboxane A2 antagonist, AA-2414 or airway hyperresponsiveness induced by ozone exposure in dogs. *Arerugi* 1991;40(1):28-36

51. Matsumoto T, Ashida Y. Inhibition of antigen-induced airway hyperresponsiveness by a thromboxane A2 receptor antagonist (AA-2414) in *Ascaris suum*-allergic dogs. *Prostaglandins* 1993;46(4):301-18
52. Nagai H, Kunimoto M, Yoshitake K, Iwama T, Arimura A, Koda A. The effect of three novel thromboxane A₂ receptor antagonists (S-1452, AA-2414, and ONO-3708) on the increase in pulmonary pressure caused by Forssman anaphylaxis in guinea pigs. *Prostaglandins, Leukotrienes, and Essential Fatty Acids* 1992; 45:233-238
53. An open-labeled, randomized, cross-over bioequivalence study of Seratrodast 80mg under fasting condition. Data on file
54. A Multi-centric, Double blind, Randomized, Comparative Clinical trial to Evaluate the Efficacy and Safety of Seratrodast 80 mg as compared to Montelukast 10 mg in the treatment of Asthma. Data on file.
55. Li X, Zhao J et al. Seratrodast in treatment of 107 patients with asthma. *Chin J New Drugs Clin Rem* 2006; 25(10): 729-733.
56. Kai-sheng Y, Ying-yun C et al. A Double-Blind Randomized Clinical Study in Multiple-Centre Comparing The Effect Of Seratrodast On Asthma With Zafirlukast. *J Jiangsu Clin Med*. 2004; Vol. 02.
57. Ishiura Y, Fujimura M, Yamamori C, Nobata K, Myou S, Kurashima K, Takegoshi T. Thromboxane antagonism and cough in chronic bronchitis. *Ann Med*. 2003; 35(2):135-9
58. Fukuoka T, Miyake S, Umino T, Inase N, Tojo N, Yoshizawa Y. The effect of Seratrodast on eosinophil cationic protein and symptoms in asthmatics. *Journal of asthma*. 2003; 40(3):257-264
59. Horiguchi T, Tachikawa S, Kondo R, Shiga M, Hirose M, Fukumoto K. Study on the usefulness of Seratrodast in the treatment of chronic pulmonary emphysema. *Arzneimittelforschung*. 2002; 52(10):764-8
60. Baba K, Sakakibara A, Yagi T, Niwa S, Wakayama H, Takagi K. Long-term observations of the clinical course after step down of corticosteroid inhalation therapy in adult chronic asthmatics: correlation with serum levels of eosinophil cationic protein. *Respirology*. 2002 Sep; 7(3):255-66
61. Tamaoki J, Kondo M, Nakata J, Nagano Y, Isono K, Nagai A. Effect of a thromboxane A2 antagonist on sputum production and its physicochemical properties in patients with mild to moderate asthma. *Chest* 2000; 118:73-79
62. Yasuo S, Tokuji U, Tomonori T, Kotaro B, Masaru O, Nobuya O. Clinical Evaluation of Seratrodast, a Thromboxane A2 Antagonist, in Patients with Perennial Allergic Rhinitis. Multi-center Comparative Double-blind Test with Terfenadine. *J Clin Ther Med*. 1991; 15(2):267-307
63. Tanaka H, Igarashi T, Saitoh T, Teramoto S, Miyazaki N, Kaneko S, Ohmichi M, Abe S. Can urinary eicosanoids be a potential predictive marker of clinical response to thromboxane A2 receptor antagonist in asthmatic patients? *Respir Med*. 1999 Dec;93(12):891-7
64. Hoshino M, Sim J, Shimizu K, Nakayama H, Koya A. Effect of AA-2414, a thromboxane A2 receptor antagonist, on airway inflammation in subjects with asthma. *J Allergy Clin Immunol*. 1999; 03:1054-61
65. Samara E, Cao G, Locke C, Granneman GR, Dean R, Killian A. Population analysis of the pharmacokinetics and pharmacodynamics of seratrodast in patients with mild to moderate asthma. *Clin Pharmacol Ther*. 1997 Oct;62(4):426-35.
66. Takeru I, Tokuji U, Kotaro B, Yasuo S, Masaru O. A Study on Efficacy and Safety of 20mg/day, 40mg/day, 80mg/day of Seratrodast, a Thromboxane A2 Receptor Antagonist on Perennial Allergic Rhinitis Patients. An Early Phase II Study. *J Clin Ther Med*. 1991; 15(2):147-182

67. Yasuo S, Tokuji U, Tomonori T, Kotaro B, Masaru O, Nobuya O. Dose Finding Study of Seratrodast, a Thromboxane A₂ Antagonist, in Patients with Perennial Allergic Rhinitis. A Multi-center, Double-blind Comparative Study. *J Clin Ther Med.* 1991; 15(2):213-245
68. Koji Y et al. Clinical Evaluation on 12-week Administration of Seratrodast, a Thromboxane A₂ Antagonist, in Perennial Allergic Rhinitis. *J Clin Ther Med.* 1991; 15(2):373-388
69. Yasuo S et al. Evaluation of Effect of Seratrodast on Nasal Passage Patency in Perennial Allergic Rhinitis by Acoustic Rhinometry. *J Clin Ther Med.* 1991; 15(2):183-211
70. Agrawal DK, Takami M, Ono S. A novel thromboxane synthetase inhibitor, DP-1904, inhibits human blood eosinophil degranulation. *Inflammation.* 1997 Feb;21(1):1-8
71. Schoonbrood DF, Lutter R, Habets FJ, Roos CM, Jansen HM, Out TA. Analysis of plasma-protein leakage and local secretion in sputum from patients with asthma and chronic obstructive pulmonary disease. *Am J Respir Crit Care Med.* 1994 Dec;150(6 Pt 1):1519-27.
72. Pizzichini E, Pizzichini MM, Efthimiadis A, Evans S, Morris MM, Squillace D, Gleich GJ, Dolovich J, Hargreave FE. Indices of airway inflammation in induced sputum: reproducibility and validity of cell and fluid-phase measurements. *Am J Respir Crit Care Med.* 1996 Aug;154(2 Pt 1):308-17.
73. Majima Y, Harada T, Shimizu T, Takeuchi K, Sakakura Y, Yasuoka S, Yoshinaga S. Effect of biochemical components on rheologic properties of nasal mucus in chronic sinusitis. *Am J Respir Crit Care Med.* 1999 Aug;160(2):421-6.
74. Basbaum CB. Regulation of secretion from serous and mucous cells in the trachea. *Ciba Found Symp.* 1984;109:4-19.
75. Wang L, Pozzato V, Turato G, Madamanchi A, Murphy TM, Chitano P. Reduced spontaneous relaxation in immature guinea pig airway smooth muscle is associated with increased prostanoid release. *Am J Physiol Lung Cell Mol Physiol.* 2008 May;294(5):L964-73
76. Tokuji U et al. Effect of Combination Therapy with Seratrodast and Mequitazine on Perennial Allergic Rhinitis. *J Clin Ther Med.* 1991; 15(2):309-337
77. Fujimoto K, Kubo K, Matsuzawa Y, Sekiguchi M. Eosinophil cationic protein levels in induced sputum correlate with the severity of bronchial asthma. *Chest.* 1997 Nov 5;112(5):1241-7.
78. Busse WW, Holgate ST. *Asthma and Rhinitis: John Wiley & Sons; 2008*
79. Ohta K, Yamaguchi M, Akiyama K, Adachi M, Ichinose M, Takahashi K, Nishimuta T, Morikawa A, Nishima S. Japanese guideline for adult asthma. *Allergol Int.* 2011 Mar;60(2):115-45.
80. Farooque SP, Arm JP, Lee TH. Lipid Mediators: Leukotrienes, Prostanoids, Lipoxins, and Platelet-activating Factor. In: *Allergy and Allergic Diseases*, Editors: Kaplan AP, Bousquet J, Holt PG, 2nd Edition, John Wiley and Sons, Ltd., Publication, 2008. pp 593-94.